

Frazionamento in Campo Flusso (FFF) e Spettrometria di massa

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Convegno Regionale SIES
Delegazione Emilia Romagna

Biopsia liquida: CHE TRAFFICO IN PERIFERIA!

Bologna

28 Febbraio – 1 Marzo 2025

Aula 1 – Complesso UniOne, Università di Bologna

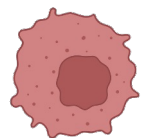
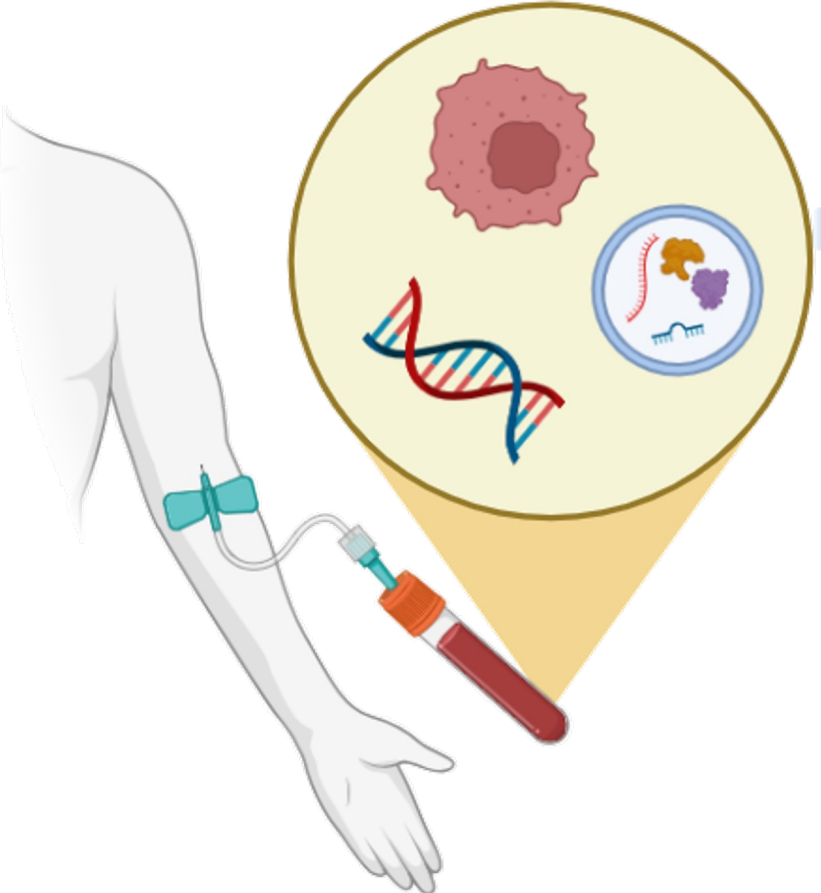


Disclosures of Valentina Marassi

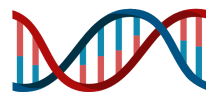
Company name	Research support	Employee	Consultant	Stockholder	Speakers bureau	Advisory board	Other
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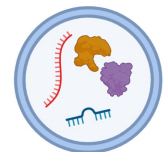
Liquid biopsy and precision medicine



Circulating tumor cells
Intact cancer cells, present as single or cluster, shed by primary and metastatic tumor

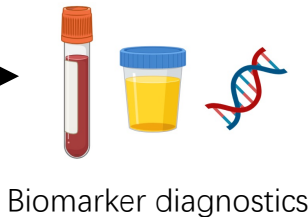


Circulating free and tumor DNA
Fragments of DNA released from cells routinely (cfDNA) and from tumor cells (ctDNA)



Extracellular vesicles
Membrane-bound, shed by cells carrying cargo (nucleic acids, proteins) and membrane markers

- Less invasive
- Early state information
- Based on cancer markers
- Requires **isolation of analytes** or **matrix-independent approaches**
- Could enhance patient-centered medicine



Biomarker diagnostics



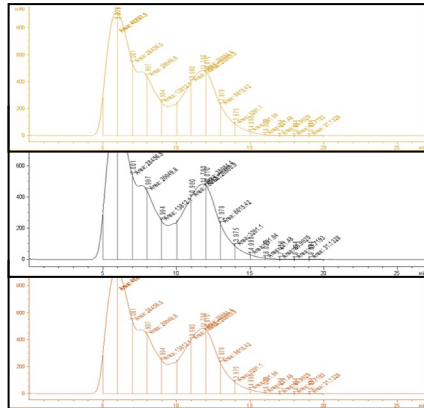
Patient-tailored therapy



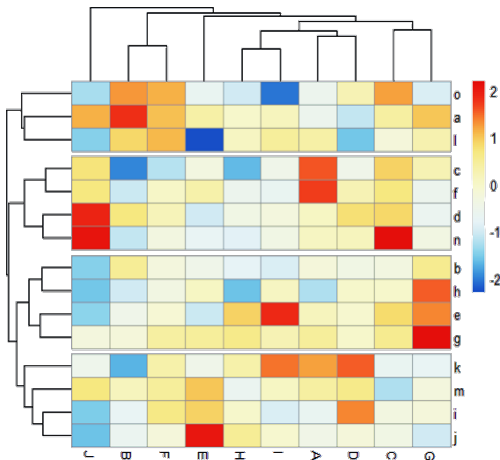
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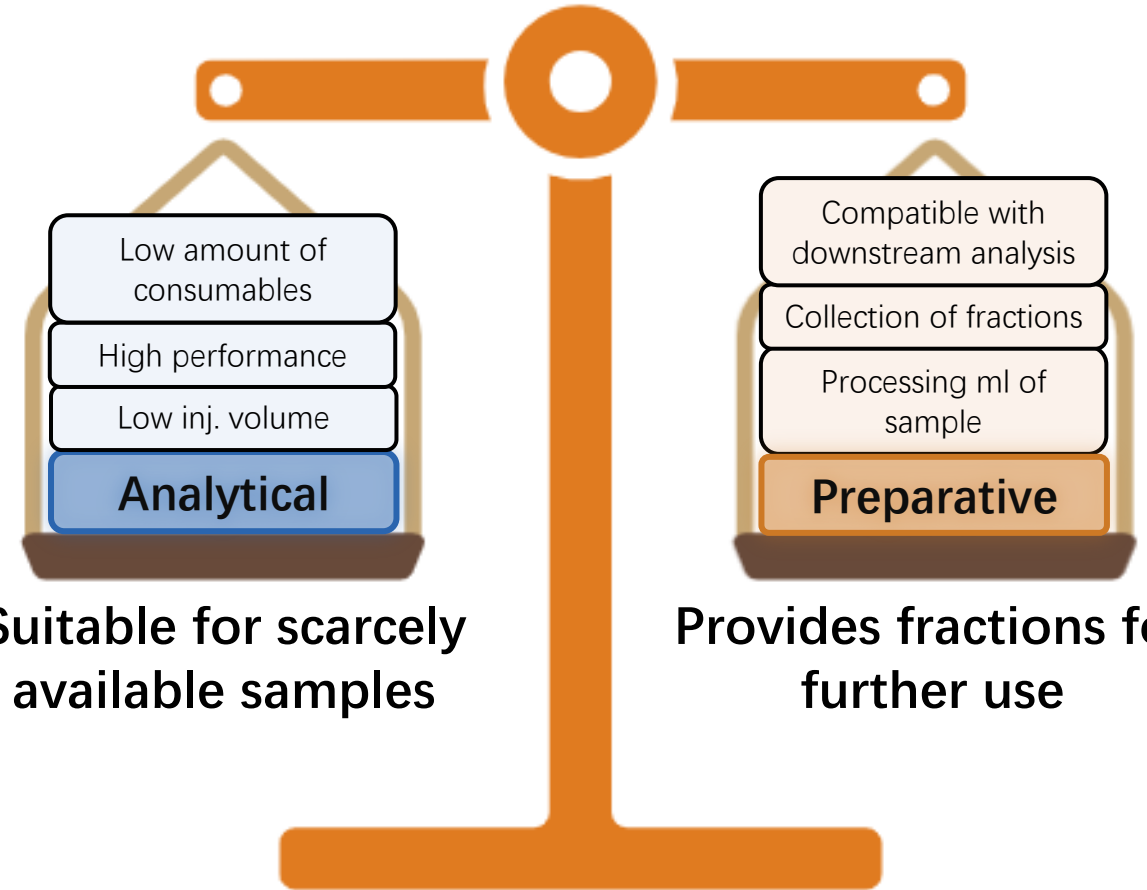
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- Simultaneous multispecies and multisize info on intact analytes



- Correlatable results
- Complete mapping
- Transversal signature



Suitable for scarcely available samples

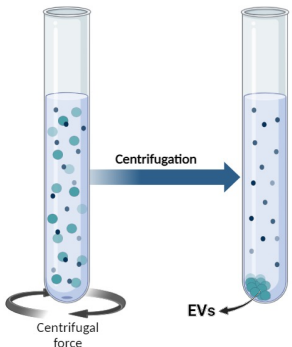
Provides fractions for further use



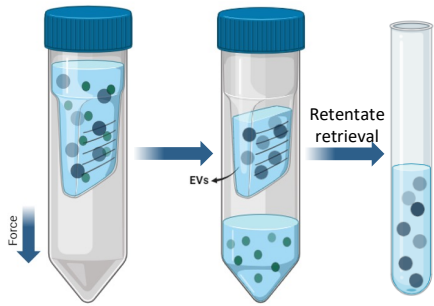
Extracellular vesicles: the challenge

➔ Characterization needs **isolation**

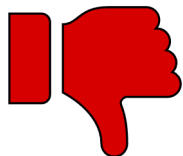
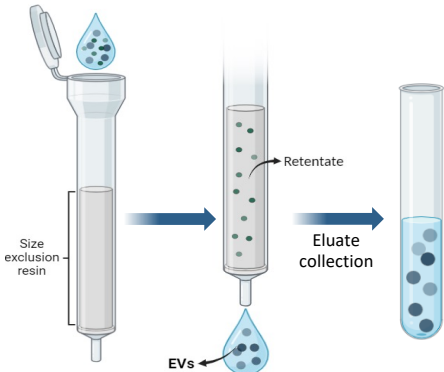
Ultracentrifugation (UC)



Ultrafiltration (UF)



Size-Exclusion Chromatography (SEC)



Limitations of conventional techniques (Ultracentrifugation, Size Exclusion, Ultrafiltration):

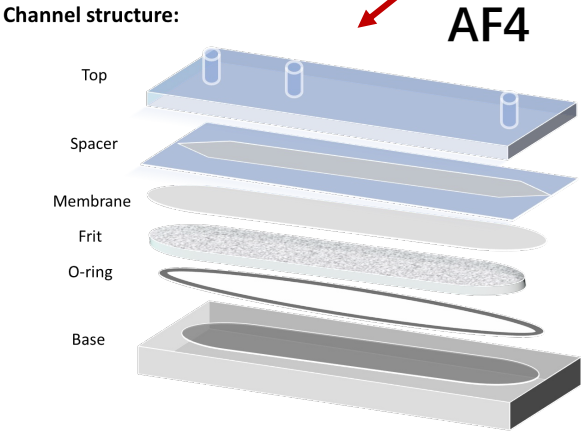
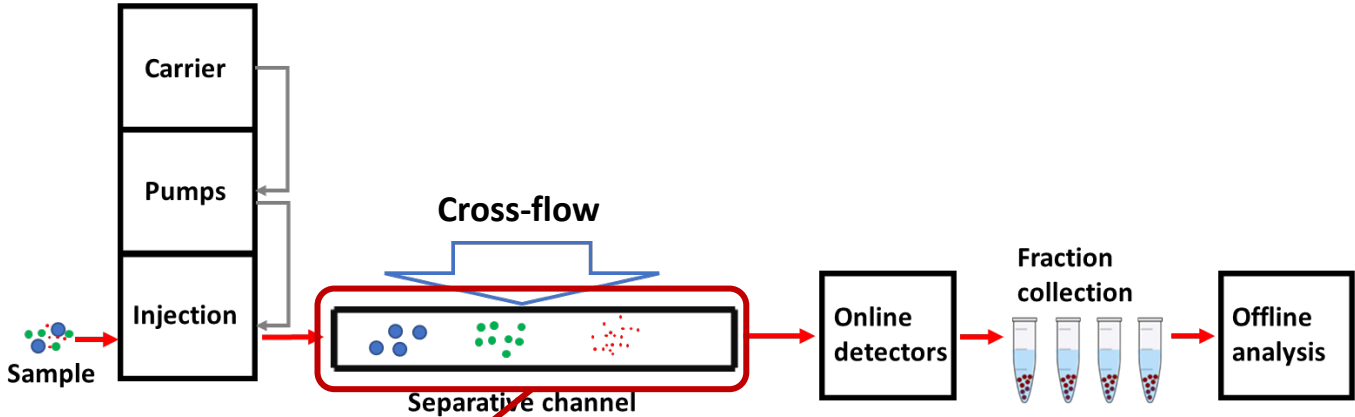
- ✗ Limited Sample flexibility (UC, UF)
- ✗ Lipoproteins co-isolation (UC, SEC)
- ✗ **EVs damage/Functionality loss** (UC, SEC, UF)
- ✗ Sample dilution (SEC)
- ✗ Sample Loss (UF, UC)
- ✗ Scale up



Adequate isolation technique required

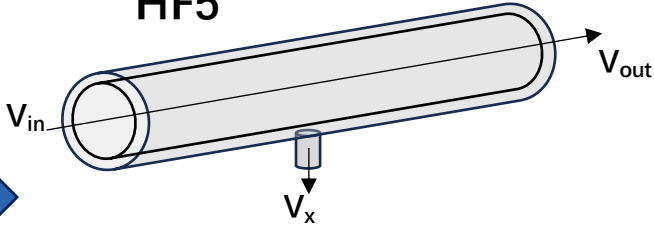


Flow Field-Flow Fractionation: the soft touch



AF4

HF5

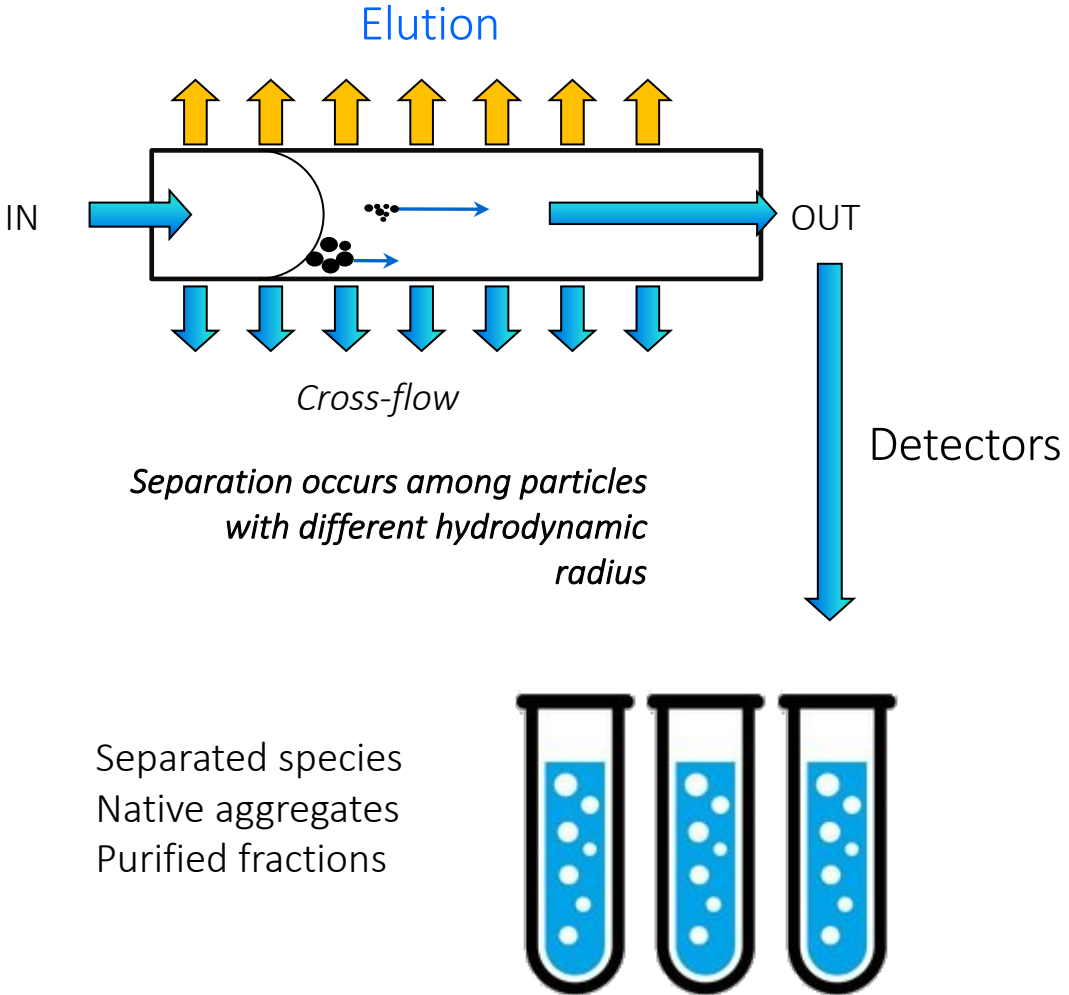
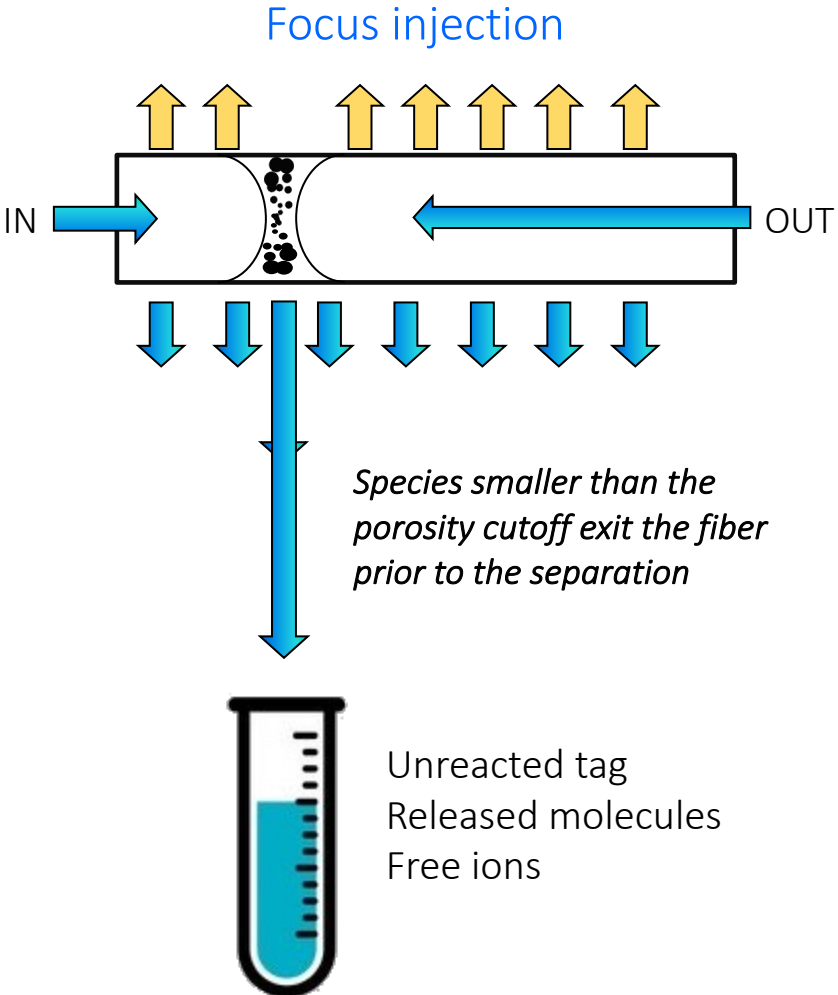


- ✓ Miniaturized version
→ minimal dilution
- ✓ Disposable cartridge

- Pseudo-chromatographic separation technique
- Hollow, porous channel
 - Sample flexibility
 - Minimal sample pretreatment
 - **Soft separation**
- Versatility on working conditions
 - Nativeness
- Analytes are size sorted
 - Hydrodynamic radius determination
- Compatible with several detectors



Method basics



The focus flow can be exploited as a "sample outlet" for simultaneous recovery of filtered and unfiltered species



AF4 Separation



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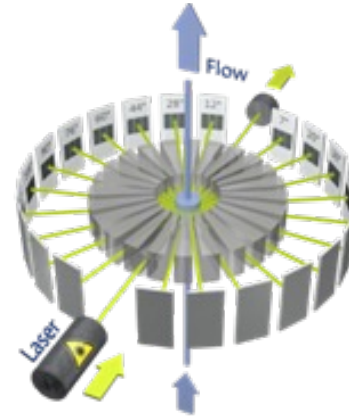
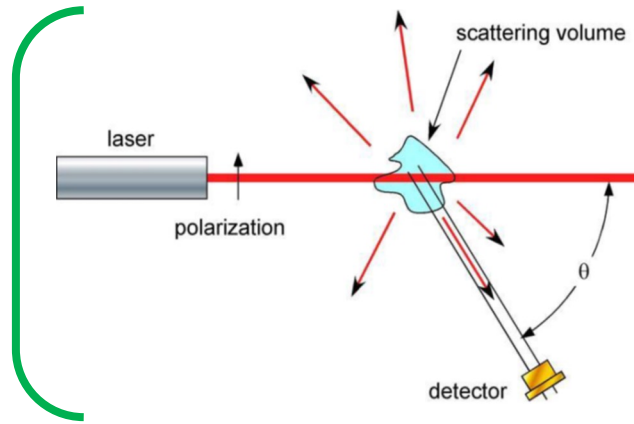
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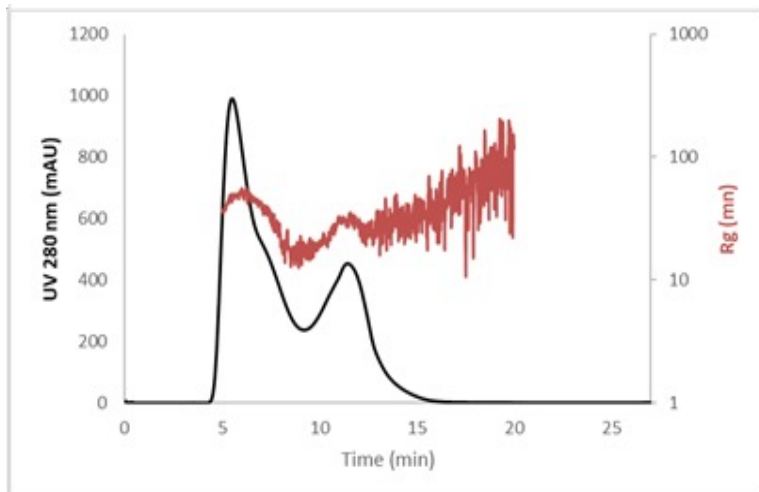
Online characterization

1) Multi Angle Light Scattering (MALS)

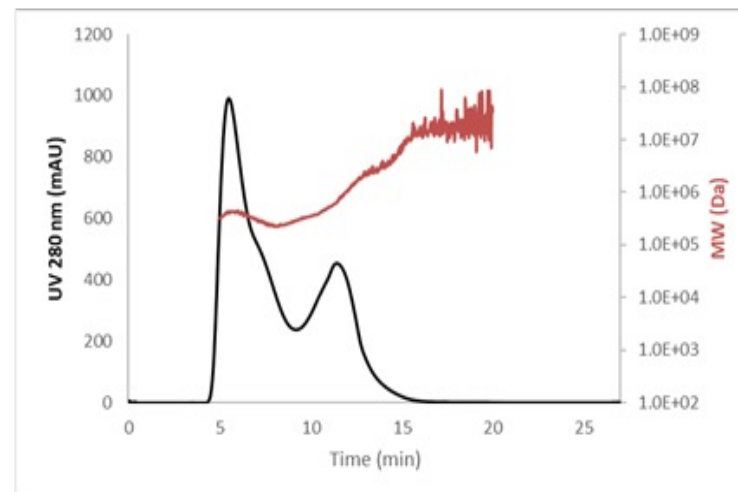


$$\frac{k^*C}{R(\theta)} = \frac{16\pi^2 n_0^2}{3\lambda_0^2} \langle r_g^2 \rangle \sin^2(\theta/2) + \frac{1}{M}$$

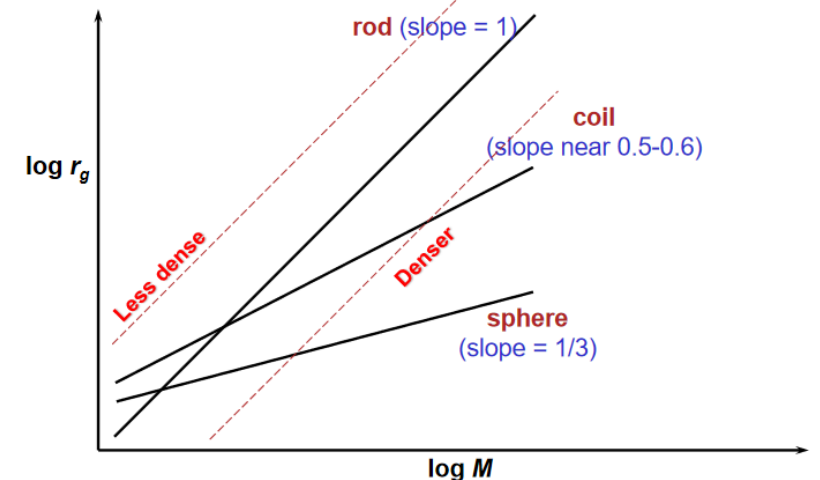
Gyration radius →
size info



Molecular weight

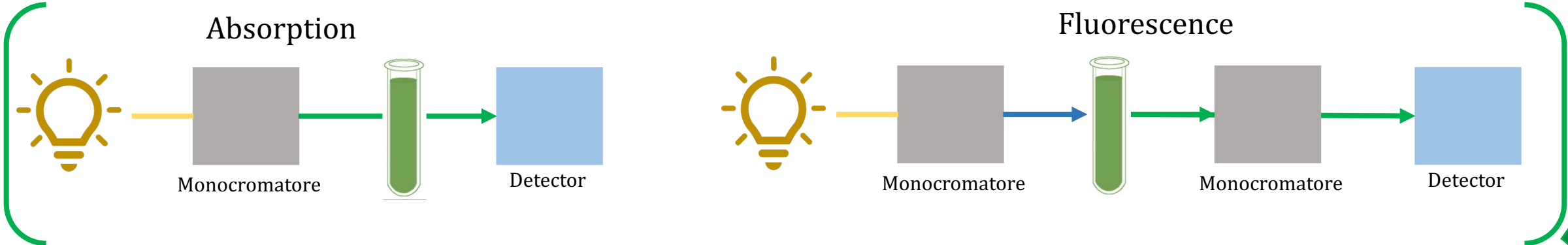


Conformation plot → shape
and morphology

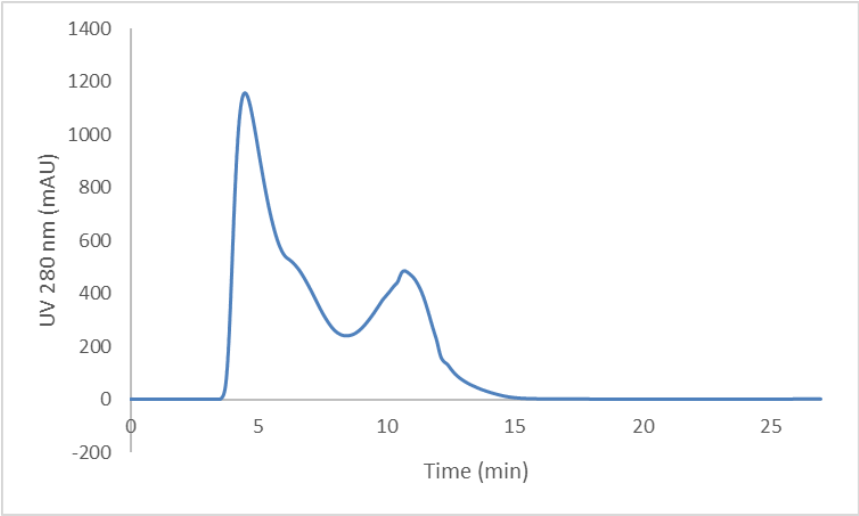
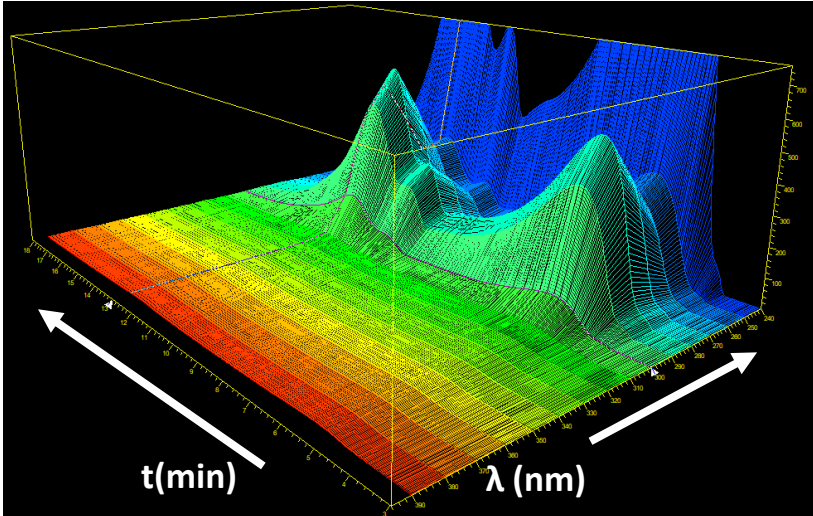


Online characterization

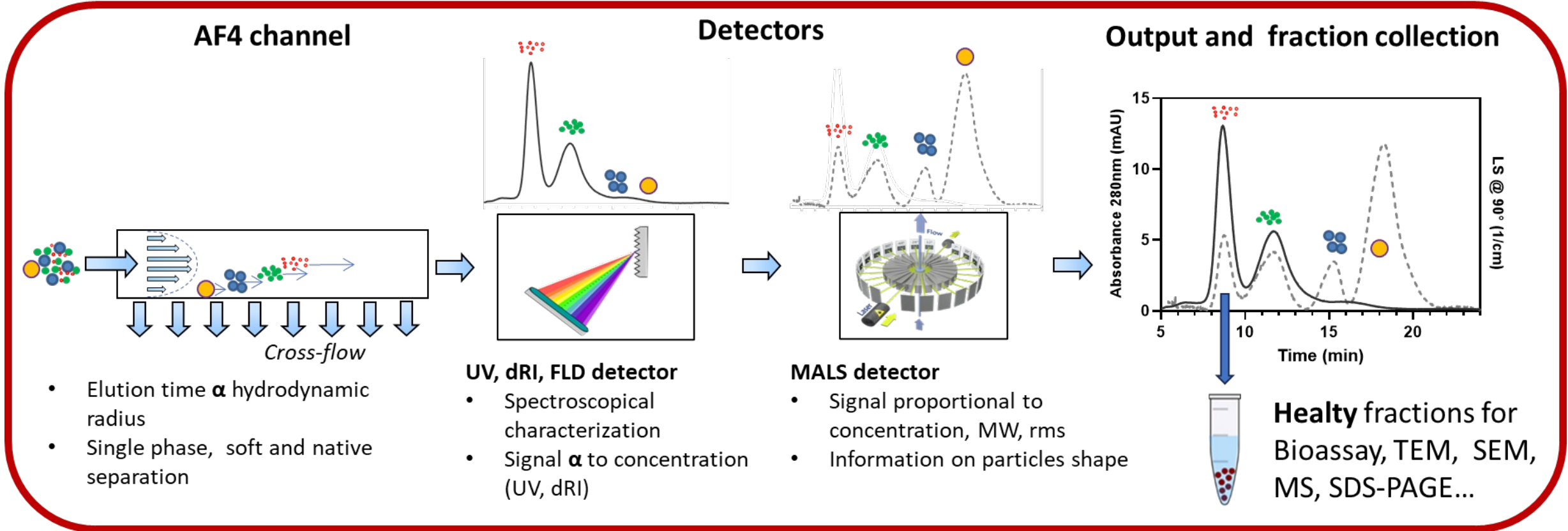
2) UV absorption and fluorescence



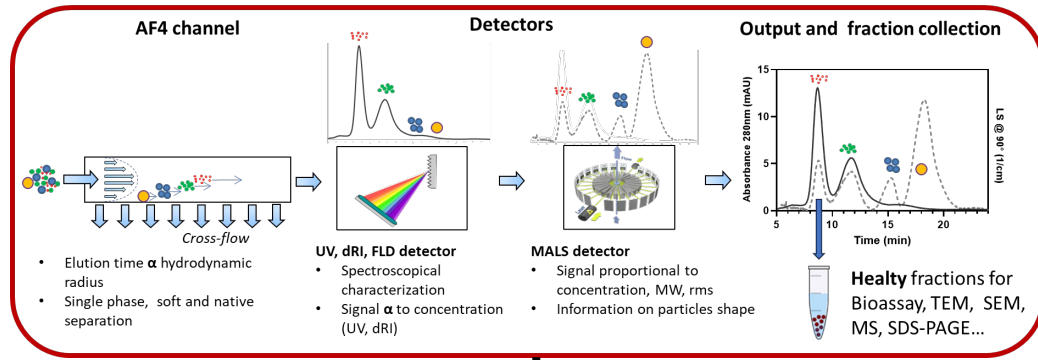
Spectral (composition) information



Platform overview



Platform overview



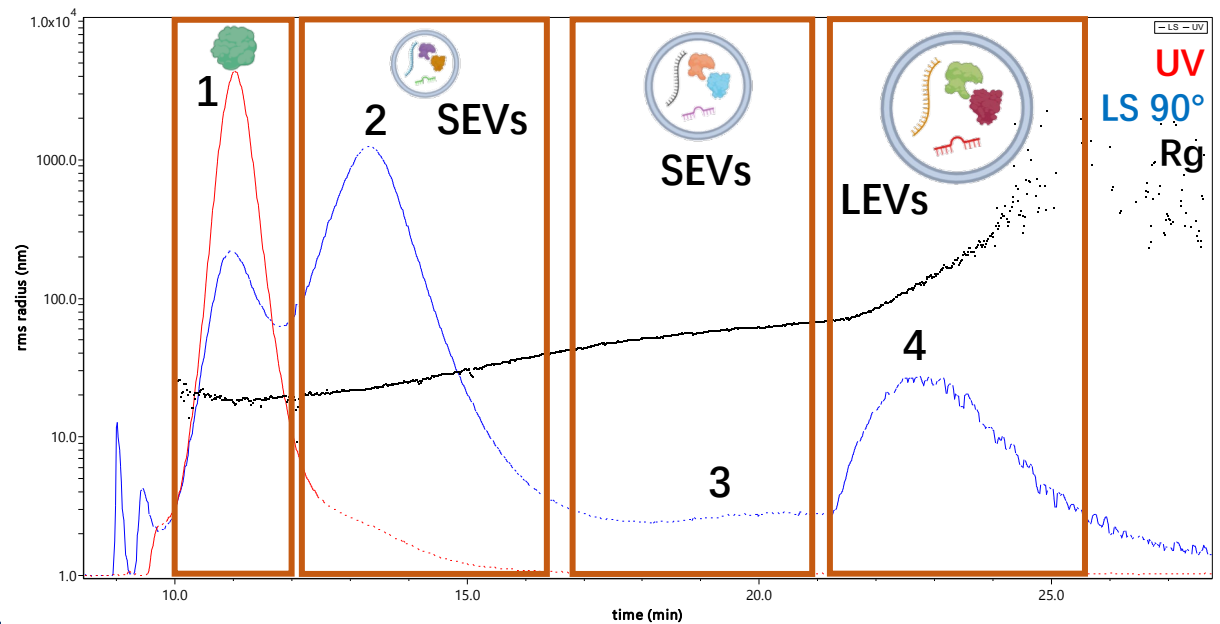
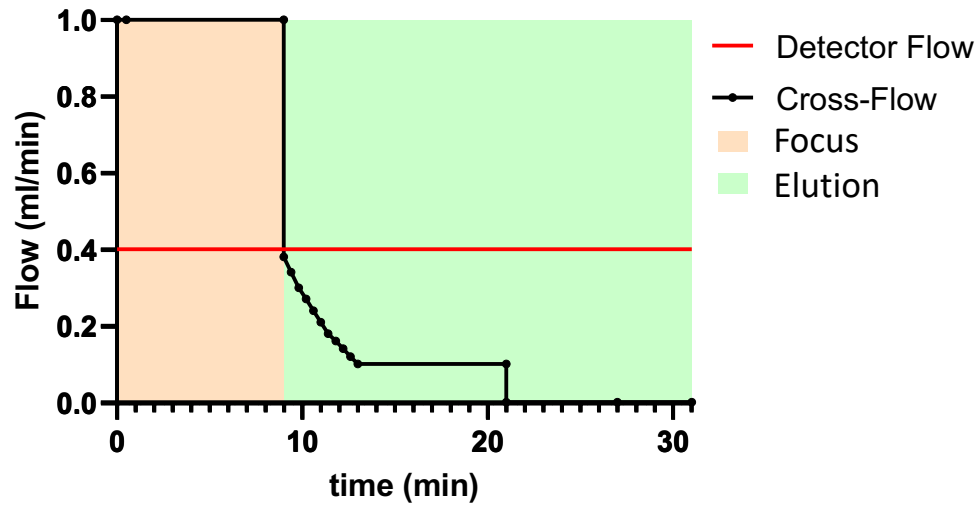
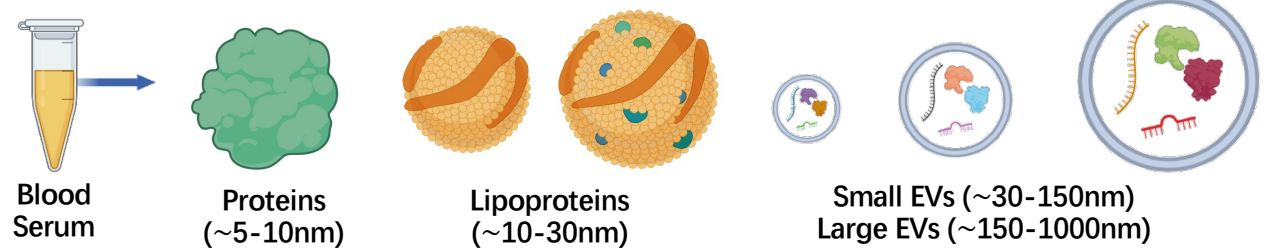
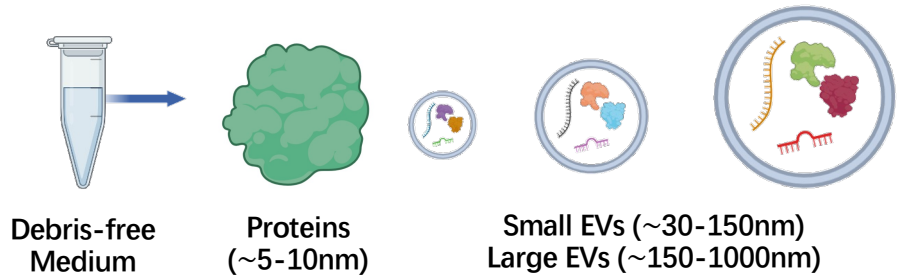
	Separation	Pretreatment	Multiparametric	Flexibility	Troughput	Destructive
AF4-MD						
Microscopy						
Chromatography						
Spectroscopy						
NTA/DLS						



Subpopulations sorting



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Mass spectrometry

To discover novel biomarkers EVs derived microRNA ,metabolites, lipids, and proteins have been investigated .

→ Among the biological molecules, exosomal proteins are most intensively investigated as biomarkers for various diseases

→ LC-MS/MS-based proteome analysis of exosomes has become the most popular fundamental tool for the identification and characterization of exosomal proteins

Bottom-Up Approach (Proteolytic Digestion-Based)

- Proteins are enzymatically digested (e.g., with trypsin) into smaller peptides before MS analysis.
- Peptides are then analyzed using liquid chromatography-MS (LC-MS/MS) and identified by database searches.

✓ Advantages:

High sensitivity and deeper proteome coverage

Well-established workflows and databases for peptide identification

Suitable for complex exosome samples

✗ Disadvantages:

Loss of information on post-translational modifications (PTMs) and protein isoforms

Requires protein digestion, increasing sample preparation complexity

Peptide reconstruction to full-length proteins can be challenging

Top-Down Approach (Intact Protein Analysis)

- Intact proteins (without digestion) are directly analyzed using high-resolution MS.
- Enables identification of proteoforms, including PTMs and sequence variations.

✓ Advantages:

Preserves complete protein structure, providing full proteoform characterization

Direct detection of PTMs and isoforms

Reduces sample processing steps compared to bottom-up

✗ Disadvantages:

Lower sensitivity, especially for complex exosome samples

Requires advanced MS instrumentation (e.g., Orbitrap, FT-ICR MS)

Challenging data analysis due to complex spectra



Steps for EV Analysis via Mass spectrometry

EV Isolation & Purification

- Techniques: **Ultracentrifugation, size-exclusion chromatography (SEC), precipitation, immunoaffinity, or field-flow fractionation (FFF)**
- Remove contaminants (e.g., proteins, lipoproteins) while preserving EV integrity

EV Lysis & Protein Extraction

- Methods: **Detergent-based lysis, ultrasonication, freeze-thaw cycles**
- Release **EV proteins** for MS analysis

Proteomic Sample Preparation

- Bottom-Up MS**: Protein digestion (e.g., trypsin) → Peptide cleanup
- Top-Down MS**: Intact protein preparation without digestion
- Enrichment for **low-abundance proteins** may be necessary

Mass Spectrometry Analysis

- Liquid Chromatography-MS (LC-MS/MS)** for peptide separation
- High-resolution MS (e.g., Orbitrap, FT-ICR, TOF-MS)** for protein/peptide identification
- DIA (Data-Independent Acquisition) or DDA (Data-Dependent Acquisition)** strategies

Data Processing & Bioinformatics

- Peptide/Protein Identification** (database search, spectral matching)
- Proteoform & PTM Analysis** (for top-down MS)

•**Quantitative Proteomics** (Label-free, SILAC, TMT/iTRAQ)

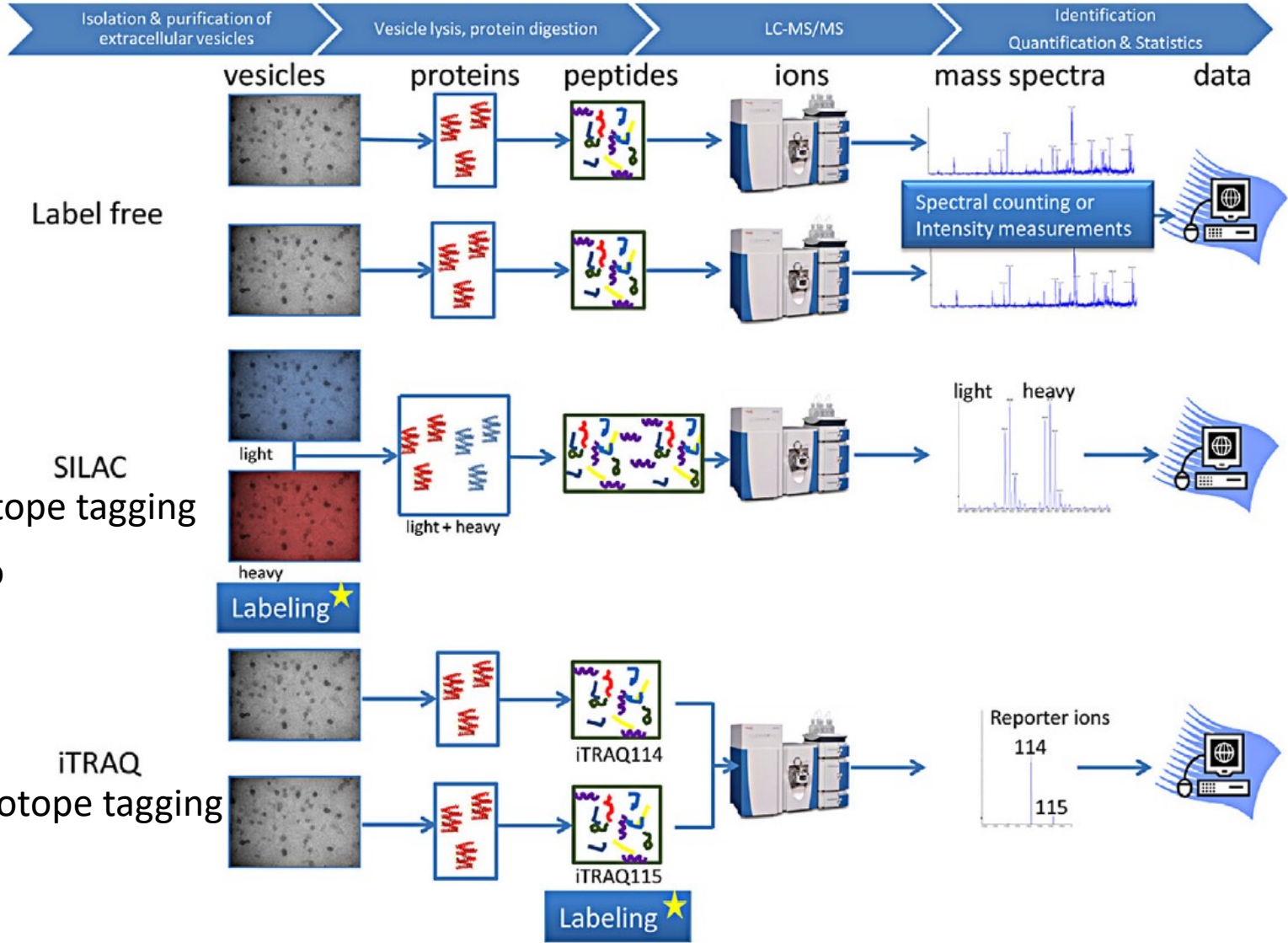
•**Pathway & biomarker discovery analysis**



Mass spectrometry

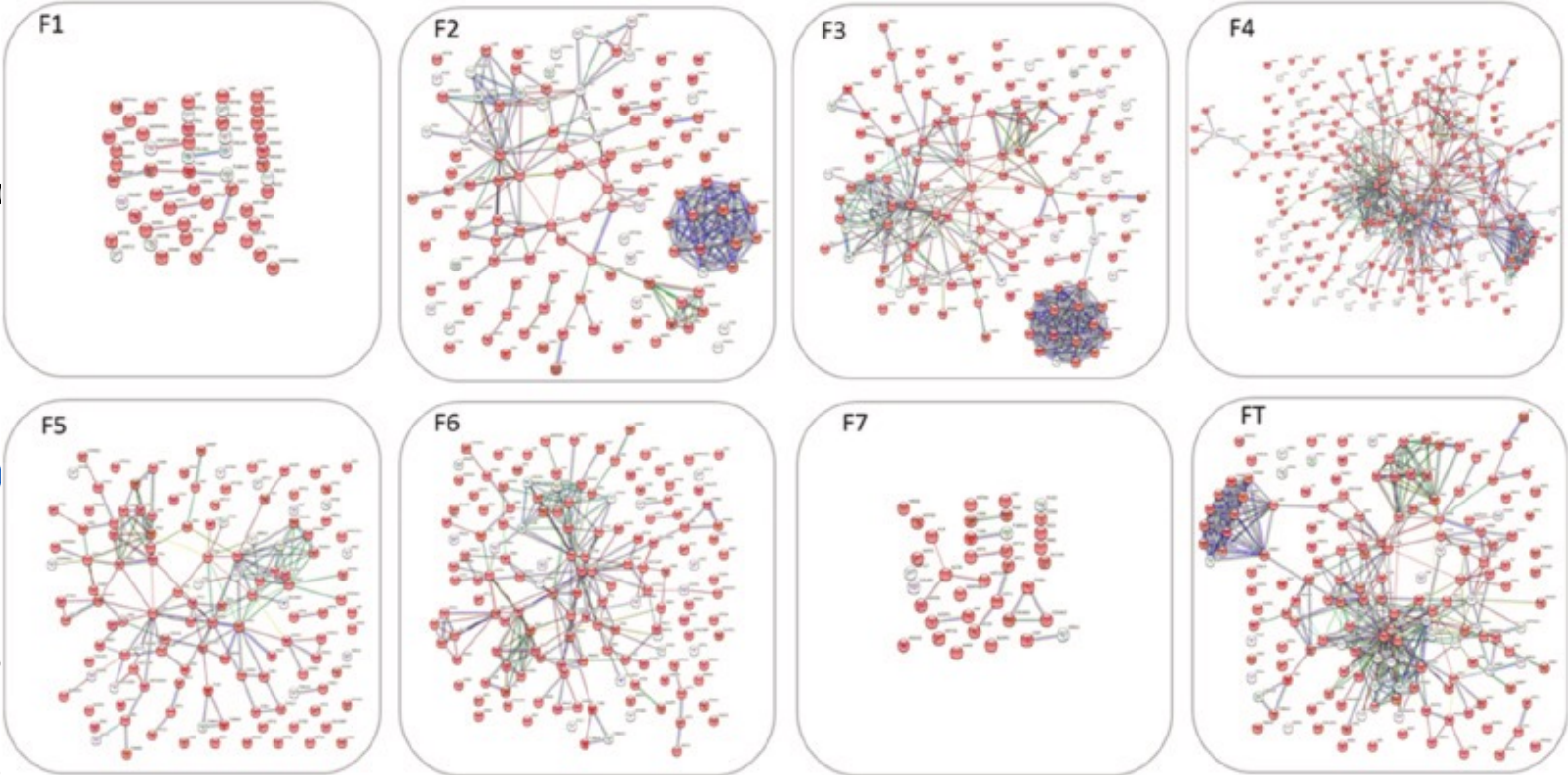
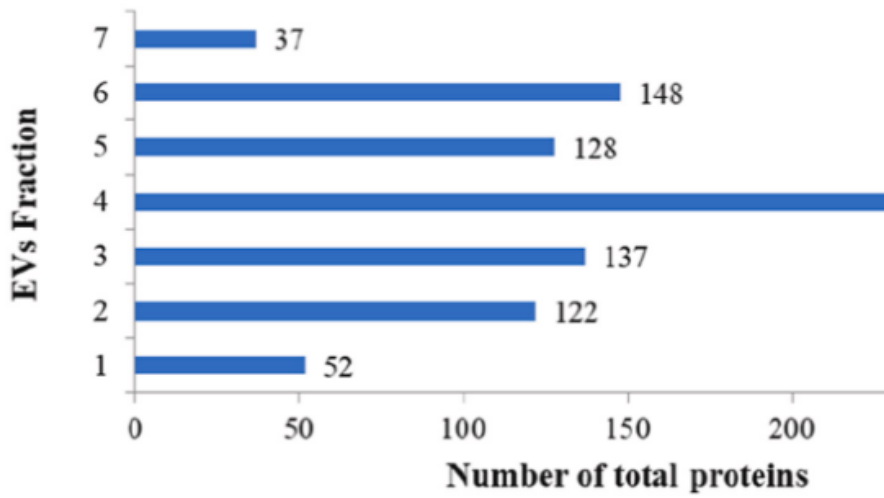
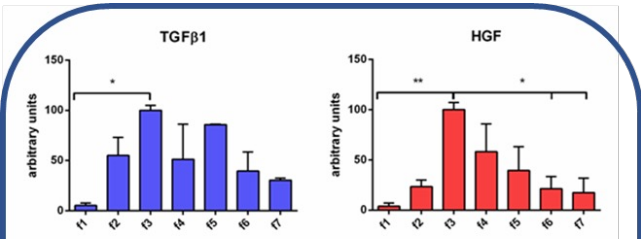
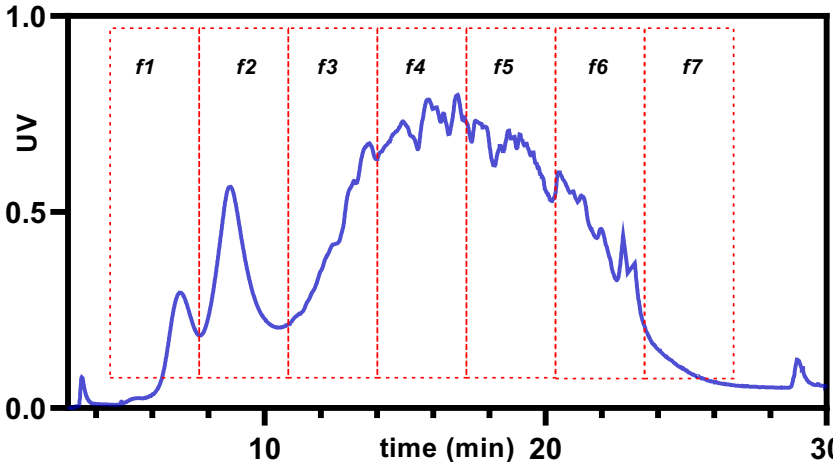
Mass spectrometry (MS), can identify and characterize molecular composition of vesicles

- MS is the major tool to assess protein composition of Evs in qualitative and quantitative proteomics approaches
- lipid and metabolite composition of vesicles might also be best assessed by MS techniques



to track EV protein cargo dynamics by labeling donor cells

FFF + Mass spectrometry



Native characterization and QC profiling of human amniotic mesenchymal stromal cell vesicular fractions for secretome-based therapy

Summary

- **Field-Flow Fractionation (FFF)** is a powerful technique for **size-based isolation** of EVs, enabling better separation from contaminants like lipoproteins and protein aggregates.
- **Mass Spectrometry (MS)** plays a crucial role in **EV characterization**, offering insights into **protein composition, biomarkers, and post-translational modifications (PTMs)**.
- **Bottom-Up vs. Top-Down MS Approaches:**
 - Bottom-up provides **higher sensitivity** but loses information on intact proteoforms.
 - Top-down preserves **full protein structure and PTMs** but faces challenges in sensitivity and data complexity.
- **FFF-MS Integration:** combining **FFF with MS** enhances the resolution and depth of EV proteomics.
- **Future Perspectives:** Further advancements in **FFF and MS methodologies, instrumentation, and data analysis** will improve **biomarker discovery** and **clinical applications of EVs**.

